

Comprehensive Invited Reviews

Approaches for regenerative healing of cutaneous wound with an emphasis on strategies activating the Wnt/ β -catenin pathway

Sehee Choi¹, Minguen Yoon¹, Kang-Yell Choi^{1,2*}

¹Department of Biotechnology, College of Life Science and Biotechnology, Yonsei University, Seoul, Korea. ²CK Biotech Inc., Rm 417, Engineering Research Park, 50 Yonsei Ro, Seodaemun-Gu, Seoul 03722, Korea.

* e-mail: kychoi@yonsei.ac.kr

A running title: Approaches for regenerative wound healing

Keywords: CXXC5, Growth factors, Stem cells, Regenerative wound healing, Wnt/ β -catenin pathway

The word count:

- Main scientific body: 5174 words (limit: 5000-7500 words)
- Abstract: 156 words (limit: < 250 words)

Significance: In adult mammals, spontaneous repair of a cutaneous wound occurs slowly and leaves a scar with skin adnexa deficiencies. To accelerate cutaneous wound healing rates and avoid scar formation, current studies have focused on regenerative therapies.

Recent Advances: Emerging therapeutics for regenerative wound healing often focus on the use of growth factors and stem cells. However, these therapeutic approaches have limited routine clinical use due to high costs and technical requirements.

Critical issue: Understanding the molecular mechanisms involved in the signaling pathways for cutaneous wound healing and neogenic synthesis of the skin components is important for identification of novel targets for development of regenerative wound healing agents.

Future Directions: The Wnt/ β -catenin pathway is a well-known key player for enhancement of the overall healing process involving tissue regeneration via crosstalk with other signaling pathways. Strategies that activate the Wnt/ β -catenin pathway via modulation of the pathway-controlling regulatory factors could provide effective therapeutic approaches for regenerative wound healing.

1.0 SCOPE AND SIGNIFICANCE

Our understanding of wound healing has increased over the past several decades along with advancements in the understanding of the molecular mechanisms involved in the process. In recent years, clinicians have been interested in the regenerative medications that increase the rates of wound healing, without scar formation. However, current therapeutic agents (e.g., growth factors) have limited use due to drawbacks such as poor efficacies, high costs, and low delivery rates. In this review, we describe distinctions between repair and regeneration in the wound healing process, then discuss current regenerative therapies and their limitations. Next, we discuss a future direction for the development of novel therapeutics that can induce regenerative wound healing by targeting Wnt/ β -catenin pathway, a key signaling pathway involving skin regeneration. Finally, we suggest an approach targeting CXXC5, a negative feedback regulator of the Wnt/ β -catenin pathway, as a safe and effective strategy for development of regenerative wound healing agents.

2.0 TRANSLATIONAL RELEVANCE

Instead of repair, which often results in scar formation, current therapeutic approaches for wound healing aim to stimulate a regenerative response that restores the wounded skin to the pre-injured state. Therefore, understanding the molecular mechanisms of the signaling pathways involving regenerative healing is important for the development of regenerative wound healing agents. The Wnt/ β -catenin pathway plays important roles in multiple wound healing processes, including cell proliferation and tissue remodeling. It also participates in stem cell activation and growth factor expression. Therefore, targeting the Wnt/ β -catenin pathway could be an ideal approach for regenerative wound healing.

3.0 CLINICAL RELEVANCE

New therapeutics have been developed subsequent to the biological and technical advances in the field of wound healing. However, current therapeutic agents are limited due to unsatisfactory efficacies, economic burdens, and adverse effects. The Wnt/ β -catenin signaling pathway is an attractive target for the treatment of many diseases related to tissue homeostasis, including wound healing. Approaches that search for Wnt/ β -catenin pathway activating compounds have been developed. A strategy that activates this

pathway via blockade of its negative feedback mechanism could be a potential approach for development of regenerative wound healing agents.

4.0 BACKGROUND and OVERVIEW

Wound healing is a complex process consisting of multiple phases (e.g., inflammation, proliferation, and remodeling) controlled by coordinated interactions among various cells, growth factors, cytokines, and chemokines.^{1,2} A failure during any of these phases results in abnormal scar or chronic wound formation. As the elderly population increases, the incidence of chronic and non-healing wounds is increasing, and the global wound care market is growing.³ Numerous wound care products and therapies have also been developed.

Conventionally, a skin wound was managed using antibiotics and wound dressings that prevent infection and ensure sufficient tissue perfusion.⁴ However, these traditional therapies cause scar, which results in functional and cosmetic impairments including increased sensitivity to ultraviolet radiation and deficiencies in skin structures (e.g., sweat glands and hair follicles). With the growing interest in higher-quality wound healing, therapeutic approaches using growth factors and stem cells have been introduced as regenerative medicines for complete recovery of the damaged tissue without a remaining scar.⁵⁻⁷ However, these approaches have limitations for routine clinical use due to drawbacks including high costs, technical difficulties, and delivery.⁸⁻¹⁰ Although a drug delivery system has been advanced, these issues remain to be resolved.¹¹

Over the past several decades, tremendous improvements have been made in understanding the relationships between signaling mechanisms and the wound healing process. The transforming growth factor beta (TGF- β) pathway, the Notch pathway, the Hedgehog pathway, and the Wnt/ β -catenin pathway are signaling pathways important for skin regeneration. The absolute requirements for these pathways during skin development have been characterized through many mammalian studies.¹² Therefore, these pathways have been suggested as targets for development of regenerative wound healing agents that contribute to the complete restoration of wounded skin.

In this review, we suggest activation of Wnt/ β -catenin pathway especially via release of the negative feedback regulation by CXXC5 as a potential therapy for acute

wound. Inhibition of CXXC5 function via blockade of its interaction with Dishevelled (DVL) enhances regenerative wound healing.¹³ This approach which activates the Wnt/ β -catenin pathway via blockade of the negative feedback mechanism is safe and results in minimal scar formation.

5.0 DISCUSSION

5.1 Repair and regeneration in wound healing

The healing of wounded skin occurs by either simple repair or regeneration. Repair implies reestablishment of the structural continuity of injured or damaged tissue, and results in scar formation with tissue dysfunction. Regeneration, on the other hand, implies replacement of the injured tissue with newly generated tissue, resulting in full restoration of tissue morphology and functionality.

Wound repair

Wound healing is a dynamic process that restores the structural and functional characteristics of damaged tissues. It includes complex cellular and biochemical interactions involving multiple types of cells, extracellular matrix, growth factors, and cytokines.^{1,2} This interactive process consists of four phases: hemostasis, inflammatory, proliferative, and remodeling (Figure 1).¹⁴ In response to injury, the requirements of each phase coordinately function to achieve tissue repair. Often, however, the repaired skin is not identical to uninjured skin and induces formation of a non-functioning mass of fibrotic tissue, or a scar.⁶

Fibrosis

Adult mammalian skin typically responds to injury by fibrotic repair. Fibrosis or scarring is attributed to deposition of excess amounts of extracellular matrix (ECM) components, such as collagen. The interposition of fibrotic tissue hinders skin adnexa formation and subsequent tissue regeneration.⁶ The inflammatory process could be involved in fibrotic healing.¹⁵ Although inflammation is crucial for protection of the body from infection by foreign organisms at the wound site, the deregulated pro-inflammatory cytokines, including interleukin (IL)- 1β and tumor necrosis factor (TNF)- α , contribute to the fibrotic process and can cause a chronic wound state or abnormal wound repair such as a hypertrophic scar and a keloid.¹⁶

Regenerative wound healing

The regenerative healing of wound is characterized by complete restoration of tissue morphology and function (Figure 2). In contrast to adult mammalian tissue, injured embryonic tissue can be completely regenerated without scarring.¹⁷⁻¹⁹

In order to determine the key factors mediating regenerative healing, it is important to understand the differences between embryonic and adult wound healing. In addition to the immature inflammatory responses mentioned above, embryonic tissue retains a relatively abundant stem cell population compared to adult tissue. Therefore, it is plausible that the restricted regenerative capacity of adult mammals is attributable to lowered population of stem cells or deficiency of proper environmental signals. Recent studies demonstrate that new tissue is generated by de-differentiation and transdifferentiation of adult cells at the edges of the wound.^{20,21} In addition, it was demonstrated that the epidermis of wounded adult mice regenerates skin adnexa, including hair and sweat glands, depending on characteristics such as wound size and status of specific signaling, e.g., the Wnt/ β -catenin pathway.²² These observations indicate that there are factors leading to an embryonic skin-like environment in the wounded skin area during the healing process of adult mammals. This could occur by de-differentiation of adult cells to a stem cell-like state or recruitment of stem/progenitor cells into the wounded region. The Wnt/ β -catenin pathway is closely related with these critical events involving activation of adult stem cells.^{23,24} Wnt/ β -catenin signaling plays important roles in determination of the fate and proliferation status of progenitor cells during embryonic development, as well as in maintenance of tissue homeostasis during the postnatal period.^{25,26} Therefore, it is worth considering the Wnt/ β -catenin pathway as a target for development of regenerative wound healing agents.

Before we discuss the role of the Wnt/ β -catenin pathway in regenerative wound healing and its underlying mechanism in detail, we will briefly review current regenerative therapies and their limitation.

5.2 Current regenerative therapies and their limitation

5.2.1 Growth factor-based therapies

- **PDGF:** PDGF participates in cell growth and angiogenesis during the wound healing process.^{27,28} PDGF comprises three isoforms: PDGF-AA, -BB, and -AB. Although PDGF-BB is a growth factor firstly approved by the US Food and Drug Administration (FDA) for application to treat chronic wounds including diabetic foot ulcers²⁹, controverting reports for its efficacy have been emerged.^{30,31}
- **EGF:** EGF induces re-epithelialization by promoting epithelial cell proliferation and migration.^{32,33} It also induces angiogenesis and tensile strength of new skin. Clinical studies found that topical application of human recombinant EGF promotes epidermal regeneration of partial thickness wounds and second-degree burns.³⁴⁻³⁶
- **TGF- β :** The main function of TGF- β is regulation of wound contraction and scar ring.^{37,38} The TGF- β family comprises three functional isoforms: TGF- β 1, TGF- β 2, and TGF- β 3. While TGF- β 1 and - β 2 facilitate fibroblast-myofibroblast differentiation and ECM deposition, they often result in fibrosis and scar formation. TGF- β 3 promotes ECM reorganization and scar reduction. Administration of Avotermin (human recombinant TGF- β 3) showed significant improvement in prevention of scar formation, but it failed to meet its endpoints in phase III clinical trials.^{39,40}
- **VEGF:** VEGF induces initiation of angiogenesis by promoting endothelial cell proliferation and migration. The VEGF family consists of VEGF-A (VEGF165), VEGF-B, VEGF-D, VEGF-E, and placental growth factor. A clinical study found that VEGF-A enhances vessel formation and improves re-epithelialization of diabetic foot wounds.^{41,42} A phase I clinical trial of the topical application of Telberim (recombinant human VEGF) found that it accelerates ulcer healing in chronic diabetic foot ulcer patients.⁴³
- **FGF:** FGF family, such as FGF-1, -2, -7, -10, and -22, have key roles during the wound healing process by promoting angiogenesis and encouraging connective t

issue cell division.^{44,45} Especially, FGF-2 (i.e., bFGF) is able to regulate collagen distribution and reduce scar formation. Clinical trials of pressure ulcer and chronic burn wound treatments found that use of FGF-2 results in acceleration of healing rates.^{46,47}

Limitations: A variety of growth factor-based therapies have been developed, but have had limited success. The development of recombinant growth factors is limited in their routine usage in clinic due to high cost and high risk. There are also many obstacles in the clinical applications because the growth factors have poor skin absorption rates and short half-lives due to their susceptibility to denaturation and proteolytic degradation.⁴⁸ Thus, they are prone to be removed by exudation before reaching the wound. In order to achieve healing, repeated administration of high growth factor concentrations is needed but, the excess dose requirement could lead to local toxicity and adverse effects such as cancer development.⁴⁹ Furthermore, growth factors are required to interact with specific surface receptors of the target cells for exhibition of their biological activities. This ligand-receptor interaction activates a series of intracellular signaling cascades followed by an expression of target genes involved in the wound healing process. However, some cells within wounds lack growth factor stimuli due to defects in molecular components (e.g., down-regulation of receptor).^{8,50,51}

5.2.2 Stem cell-based therapies

- **Mesenchymal stem cells:** Mesenchymal stem cells (MSCs) have self-renewal and multipotent differentiation characteristics.^{52,53} They can release a variety of paracrine factors that enhance wound healing (e.g., PDGF-BB, VEGF, and bFGF) and subsequently promote angiogenesis.^{54,55} MSCs have anti-bacterial and anti-inflammatory properties via the secretion of anti-microbial factors and anti-inflammatory cytokines, respectively.⁵⁶ One clinical study found that chronic skin wound healing was improved by local application of bone marrow- and adipose-derived MSCs.⁵⁷
- **Hematopoietic stem cells:** Hematopoietic stem cells (HSCs) are self-renewing cells present in the bone marrow. When transplanted into mice, HSCs can differ

entiate into follicular epithelial cells, sebaceous gland cells, and epidermal keratinocytes.⁵⁸ They also promote angiogenesis. Topical application of HSCs led to improvement in wound closure rates of full-thickness excisional wounds in diabetic mice.⁵⁹

- **Epithelial stem cells:** Epithelial stem cells (EpSCs) are quiescent cells, but they self-renew and differentiate into at least one cell type. EpSCs have an important role in stratified epidermal regeneration.⁶⁰ They also participate in hair follicle neogenesis in the wound dermis. Transplantation of a bioengineered dermis containing EpSCs into acute wounds induces skin reconstruction and hair formation in goats.⁶¹ Injection of EpSCs enhances vascularization, elastin content, and follicle-like structures when compared with the control group.⁶²

Limitations: Although stem cell therapy is a fast-growing field in regenerative medicine, many issues remain to be resolved (e.g., low safety, high cost, difficulty in administration and quality control) before the routine clinical usage. There is currently no FDA-approved stem cell therapy for wound treatment despite numerous attempts. The age of the transplanted cells and the local microenvironment of the injured skin also need to be considered for therapeutic application of stem cells.⁶³ For example, MSCs derived from old mice rather inhibited wound healing in diabetic mice.⁶⁴ A favorable environment, including a sufficient blood supply, receptor, and presence of biological molecules is necessary for effective use of stem cells as therapeutics for wound healing. These matters, together with the safety and quality control issues, make the clinical use of stem cell-based therapeutics challenging.

5.3 Future direction for the development of regenerative therapeutics

5.3.1 Signaling pathway and wound healing

In order to overcome current limitations of regenerative therapeutics and to discover new therapies, it is necessary to understand pathways involved in wound healing. Growth factor therapies eventually exert their biological activities through the downstream signaling pathway after interaction with specific surface receptors.

The function of adult stem cells residing within tissues are modulated and reprogrammed by their microenvironment, especially molecular pathways.^{65,66}

Moreover, many mammalian studies have elucidated that flawless regeneration of embryonic skin wound absolutely depends on activity of signaling pathways which are important for complete restoration of adult wound skin.¹² TGF β , Notch, Hedgehog, and Wnt/ β -catenin pathways are major players for regenerative wound healing.

The TGF- β pathway is differentially involved in the regulation of healing rate depending on the isoforms.¹² TGF- β 1 functions as a fibrosis-stimulating factor but TGF- β 3 regulates anti-scarring activity.^{67,68} Members of the TGF- β superfamily participate in the development of skin or its adnexa, such as hair follicles.^{69,70} The Notch pathway regulates epidermal cell differentiation during stages of adult and embryonic development.⁷¹ This pathway also has important roles in the maintenance of skin homeostasis and promotion of angiogenesis.⁷²⁻⁷⁴ The Hedgehog pathway is involved in skin morphogenesis and angiogenesis. The Hedgehog pathway modulates dermal repair and wound vascularization during the wound healing process.^{75,76} The Wnt/ β -catenin pathway plays a role in adult tissue regeneration⁷⁷⁻⁷⁹, and participates in multiple steps of the wound healing process together with activation of stem cells residing within skin.²⁴ Proper regulation of the Wnt/ β -catenin pathway is crucial for flawless and complete regeneration of wounded skin. Therefore, we are going to focus on the Wnt/ β -catenin pathway as a target for development of regenerative wound healing agents in the following sections of this review. Initially, small molecules or natural products which activate the Wnt/ β -catenin signaling will be introduced because those are easier to manufacture than growth factors or stem cells. In the later part, we will focus on a new strategy for the regenerative wound healing targeting CXXC-type zinc finger protein 5 (CXXC5), a negative feedback regulator of the Wnt/ β -catenin pathway functioning via interaction with DVL.^{13,80} This approach blocking the CXXC5–DVL interaction for activation of the Wnt/ β -catenin signaling could be an effective and safe way for regenerative wound healing.

5.3.2 Regulation of the Wnt/ β -catenin pathway

The Wnt/ β -catenin pathway has essential roles in numerous biological processes including cell proliferation, differentiation, and migration. It is well-known for its role in stem cell self-renewal and differentiation during normal tissue homeostasis and tissue regeneration after injury.^{78,79,81,82} The Wnt/ β -catenin pathway has recently been characterized as a key modulator of cutaneous wound healing; Wnt/ β -catenin signaling is up-regulated by wounding and is involved in the overall stages of the healing process.²⁴ Therefore, activation of Wnt/ β -catenin pathway is an attractive strategy for cutaneous wound healing. The rate of development of wound healing agents that activate the Wnt/ β -catenin pathway has increased in recent years.

Wnt/ β -catenin signaling pathway

The hallmark of Wnt/ β -catenin signaling activation is stabilization and nuclear translocation of β -catenin. The stability of β -catenin is regulated by formation of a destruction complex consisting of Axin, adenomatous polyposis coli (APC), glycogen synthase kinase-3 (GSK-3), and casein kinase 1 (CK1) in the cytoplasm (Figure 3).⁸³⁻⁸⁵ In the absence of Wnt stimuli, β -catenin is subjected to priming phosphorylation by CK1 and subsequent phosphorylation by GSK-3 in the destruction complex (Figure 3). The recruitment of β -TrCP, a E3 ubiquitin ligase, to the phosphorylated β -catenin results in proteasomal degradation via polyubiquitination. When extracellular Wnt ligands bind to the Frizzled receptor and LRP5/6 co-receptor complex, recruitment of the downstream signal mediators, such as Dishevelled (DVL) and Axin, is triggered and results in the dissociation of the destruction complex. Ultimately, the β -catenin is freed from the complex, accumulates in the cytoplasm, and is then translocated into the nucleus. In the nucleus, β -catenin binds with the T-cell factors/lymphoid enhancing factors (TCFs/LEFs) for target gene expression. More than 100 Wnt/ β -catenin target genes have been identified.⁷⁹ Many of these genes (e.g., *Axin2*, *Collagen I*, *Collagen III*, *EGFR*, *Endothelin-1*, *Fibronectin*, *Keratin-14*, *Lgr5*, *VEGF*, and *WISP1*) have roles in cutaneous wound healing (Figure 4 and Table 1).

The roles of Wnt/ β -catenin signaling in the wound healing process

Many target genes that are transcriptionally induced by activation of the Wnt/ β -catenin pathway mediate various functions during the wound repair process (e.g., inducing structural construction of the dermis and epidermis and promoting angiogenesis) (Table 2). Moreover, the Wnt/ β -catenin pathway plays key roles in regenerative wound healing (e.g., inducing formation of skin adnexa, such as hair) by promoting the activation of the stem cells.²⁴

- The profile of Wnt/ β -catenin signaling activity during the wound healing process

The Wnt/ β -catenin signaling pathway is activated in the dermis of the wound bed soon after a skin injury. This activation is quick and spatially restricted within the wound site. A study displayed that the change of oxygen tension occurring within minutes of skin damage can trigger activation of the Wnt/ β -catenin pathway through hypoxia-inducible factor (HIF)-1 α .⁸⁶ During the proliferative phase of wound healing, Wnt/ β -catenin signaling activity is highly increased in mesenchymal cells. For example, murine dermal fibroblast cultures exhibit increment in β -catenin protein levels and TCF/LEF-mediated transcriptional activity during proliferation.⁸⁷ In human wound samples, the levels of β -catenin and the expression of its target genes (e.g., fibronectin and MMP7) are increased during the dermal proliferative phase.⁸⁸

- The roles of Wnt/ β -catenin pathway in wound repair

Up-regulation of Wnt/ β -catenin signaling promotes proliferation and migration of dermal fibroblasts, making them differentiate into myofibroblasts. This process helps to reduce the surface area of the developing scar.^{87,89} The activated Wnt/ β -catenin signaling not only facilitates migration and differentiation of keratinocytes in the epidermis, but it also promotes angiogenesis, follicle regeneration, and epithelial remodeling, which directly enhances cutaneous wound healing.^{24,90}

- ***The roles of Wnt/ β -catenin pathway in stem cell activation***

Typically, stem cells are harbored in most adult tissues. However, they easily lose their self-renewal capability in response to stress or aging. The Wnt/ β -catenin pathway plays roles in tissue-residing, stem cell activation, and migration to the wound bed in the basal epidermis for regeneration of damaged tissue.⁹¹ In response to the Wnt/ β -catenin signaling, TCF/LCF complex modulates the fates of lineages of multipotent stem cells in the skin.⁹² For example, Wnt/ β -catenin is involved in activation of epidermal stem cells (ESCs), the major source for replenishment of lost cells in the process of wound healing.⁹³ The elevated Wnt/ β -catenin signaling activity not only enhances proliferation of quiescent ESCs, but it also promotes differentiation of ESCs into keratinocytes.⁹⁴ Furthermore, elevation of β -catenin activity significantly promotes neogenesis of hair follicles, representing a fully functional inter-follicular epidermis in adult mice.^{22,95,96} By contrast, inhibition of Wnt/ β -catenin signaling during skin wounding hinders formation of epithelial adnexa, including hair and sweat glands, resulting in scarring. These findings indicate that the Wnt/ β -catenin pathway can trigger regeneration of wounded skin by serving as a niche signal for activation of skin stem cells.⁹⁷ The Wnt/ β -catenin signaling target gene product, Axin2 and Lgr5, are well-known markers for the self-renewing stem cells in tissues including the skin and the hair follicle.^{77,98}

- ***The roles of Wnt/ β -catenin pathway in hyaluronic acid synthesis***

In wound healing, the main difference between repair and regeneration comes from ECM content. During the healing process, granulation tissue, which is formed at the wound site, is characterized by the proliferation of fibroblast that produce ECM components including fibronectin, collagen-III, elastin, and hyaluronic acid (HA).⁹⁹ In the final stage of wound repair, the granulation tissue is converted to fibrotic scar tissue as collagen-III is replaced by collagen-I forming collagen fibers. The alignment of excessive collagen fibers in the dermis results in an inelastic collagen scar. In the process of regenerative

healing, on the other hand, much higher amount of HA is present in ECM compared with the repair process, and thus it reduces collagen deposition and subsequent fibrotic scar formation.¹⁰⁰ Wnt3a treatment up-regulates genes involved in HA synthesis in fibroblasts.¹⁰¹ Furthermore, HA interacts with CD44, a well-known Wnt/ β -catenin signaling-target, in order to exert its function in wound healing process including enhancement of cell migration toward wound sites and promotion of angiogenesis as well as direct enhancement of tissue regeneration.¹⁰²⁻¹⁰⁴ Therefore, Wnt/ β -catenin pathway not only induce HA synthesis, but also regulate biological function of HA for regenerative wound healing.

- ***Crosstalk of Wnt/ β -catenin pathway with other signaling pathways***

The Wnt/ β -catenin pathway cooperates with other signaling pathways during the wound healing process. It interacts with the TGF- β /Smad pathway, which is a major signaling pathway involved in cutaneous wound healing and dermal fibrosis. TGF- β signaling is transiently activated after a skin injury; β -catenin level is then increased via the inhibition of GSK-3 β activity or DKK-1 expression.¹⁰⁵⁻¹⁰⁷ The proliferation of fibroblast and its differentiation into myofibroblasts via activation of TGF- β signaling occurs in a β -catenin-dependent manner. These results indicate that the Wnt/ β -catenin pathway is a mediator of TGF- β /Smad signaling-induced wound healing. Synergistic activation through mutual interaction of the Wnt/ β -catenin and Notch pathways improves wound healing and inhibits scar formation by promoting embryonic stem cell proliferation, keratinocyte differentiation and migration, and follicle regeneration.⁹⁴ To reconstitute skin adnexa and obtain complete healing, Wnt/ β -catenin signaling also facilitates hair follicle regeneration in wounded skin through formation of a positive feedback loop with FGF-9 signaling.¹⁰⁸

Therapeutic potential of Wnt activators for regenerative wound healing

Considering the roles of the Wnt/ β -catenin pathway during wound healing, it has recently been used as a target for the development of wound healing agents.

Studies have examined on small molecules and natural products that activate the Wnt/ β -catenin pathway as potential therapeutics for diverse diseases.^{109,110} Some of these are under development for the treatment of skin wounds (Figure 5 and Table 3).

- ***Lithium chloride:*** Lithium chloride (LiCl) is well-known GSK3 inhibitor and has a capability for activation of Wnt/ β -catenin pathway.¹¹¹ Topical application of LiCl to the wounded skin of rats induced enhancement of the wound closure rate with elevated β -catenin level.⁹⁴ Moreover, thickness of the neoformative epidermis layer and formation of hair follicle structures and sebaceous gland were increased in skin tissues of rats by topically applied LiCl.
- ***Valproic acid:*** The small-molecule valproic acid (VPA) is known to activate the Wnt/ β -catenin pathway by inhibiting GSK-3 β .¹¹² Furthermore, VPA enhances wound healing through promotion of neo-epidermis formation, fibroblast-myofibroblast transition, and cellular proliferation. One study found that when mice were treated with VPA, full-thickness wound sizes were markedly reduced, and healing rates increased. VPA also induces the expression of stem cell markers (e.g., CD34) involved in neo-vascularization.
- ***Lucidone:*** Lucidone, a naturally occurring cyclopentenone isolated from the dried fruits of *Lindera erythrocarpa*, was reported to increase β -catenin level through GSK3 β -dependent pathway and enhance wound healing both *in vitro* and *in vivo* models.¹¹³ Lucidone not only promoted proliferation and migration in both keratinocyte and fibroblast cells, but also triggered expression of angiogenesis markers in endothelial cells. The healing rate of punched wounds on mice was accelerated by the topical application of lucidone.
- ***Polygonum aviculare L.:*** *Polygonum aviculare L.* extract was screened out as a natural product that activates the Wnt/ β -catenin pathway.¹¹⁴ At the cell level, *P. aviculare L* extract promoted migration of both keratinocytes and fibroblasts

¹¹⁵ Treatment with *P. aviculare* L. extract accelerated the healing rates of full-thickness dorsal wounds in mice, compared with a control group. Active ingredients of *P. aviculare* L. extract (e.g., quercitrin hydrate, caffeic acid, and rutin) have been characterized as components that activate the Wnt/ β -catenin pathway and enhance keratinocyte migration.

5.3.3 CXXC5, a negative feedback regulator of the Wnt/ β -catenin pathway, suppresses the wound healing process

Activation of the Wnt/ β -catenin pathway is an ideal strategy for regenerative wound healing. However, development of therapeutics that activate this pathway is limited due to the presence of its own negative regulation systems. Inhibitory factors of the Wnt/ β -catenin pathway disrupt skin wound healing. For example, Dickkopf-1 (DKK-1), a secreted Wnt antagonist functioning via interaction with LRP5/6 receptor, hinders proliferation of dermal fibroblasts as revealed by both *in vitro* and *in vivo* system.¹¹⁶ Consistent with these results, intradermal injection of small interfering RNA (siRNA) for *DKK-1* enhances dermal fibroblast functions. Another secreted Wnt antagonist, Frizzled-related protein-1 (sFRP-1), suppresses cell proliferation and ECM production in keloid fibroblasts by inhibiting Wnt/ β -catenin signaling through interacting with either Wnt or Frizzled.¹¹⁷ A mouse model revealed that injection of a neutralizing antibody against sFRP-1 into the palatal wound edge promotes the healing of wounded skin.¹¹⁸

The function of a negative feedback regulator of the Wnt/ β -catenin pathway, CXXC5, is determined by its subcellular location that depends on the tissue type and the cell's physiological status. Cytosolic CXXC5 plays a role as a Wnt/ β -catenin signaling inhibitor, whereas nuclear CXXC5 functions as a transcription factor.¹¹⁹⁻¹²³

By binding DVL, cytosolic CXXC5 has a variety of pathophysiological roles by inhibiting the Wnt/ β -catenin signaling pathway (Figure 6).^{13,80,124,125} CXXC5 can be transcriptionally induced by the Wnt/ β -catenin signaling itself or under a variety of pathophysiological status (e.g., alopecia, osteoporosis, wound formation, and

termination of height growth at puberty), and these pathological aberrancies can be restored in *CXXC5* knock out mice of the disease model systems.^{13,80,124,125}

Taken together, these findings indicate that *CXXC5* can be a therapeutic target for diseases caused by suppression of Wnt/ β -catenin signaling. The importance of targeting cytosolic *CXXC5* function in wound healing was supported by enhanced cutaneous wound healing in mice treated with the protein transduction domain-fused DVL-binding motif (PTD-DBM) peptide, which blocks *CXXC5*–DVL protein-protein interactions (PPI).¹³

A role of CXXC5 during wound healing

In melanoma patients, β -catenin level was gradually increased, especially during the late inflammatory and early proliferative stages, and then decreased during the remodeling stage (Figure 7).¹³ In contrast, *CXXC5* level was declined during the early proliferative stage after surgery but then rose again, which shows the opposite patterns of those of β -catenin in the same wounded area. The inhibitory role of *CXXC5* in wound healing was revealed by the increment of the wound closure rate in *Cxxc5*^{-/-} mice.¹³ Myofibroblast differentiation and collagen production is inhibited by *CXXC5* overexpression in human dermal fibroblasts. Both *in vitro* and *in vivo* studies found that the inhibitory roles of *CXXC5* during the wound healing process are exerted by suppression of Wnt/ β -catenin signaling via its interaction with DVL in the cytosol.¹³ Taken together, these findings indicate that the inhibition of *CXXC5* function, especially its cytosolic role related to Wnt/ β -catenin signaling inhibition, may be a new strategy for development of wound healing agents.

Effects of PTD-DBM, an interfering peptide against CXXC5–DVL interaction, on wound healing

A PTD-DBM peptide, that contains the sequence of *CXXC5* binding to DVL and activates Wnt/ β -catenin signaling via interference of the *CXXC5*–DVL interaction, was developed for practical application (Figure 8A).⁸⁰ The PTD-DBM peptide effectively promotes cell migration *in vitro* via activation of Wnt/ β -catenin signaling.¹³ Topical application of the PTD-DBM peptide significantly accelerates the wound closure rate. Increment in wound healing-related markers and critical

deposition of collagen occur during the healing of full-thickness wounds on the backs of mice (Figure 8B). In addition to enhancement of wound healing, induction of alkaline phosphatase (ALP) in PTD-DBM peptide-treated mouse wounds indicate that PTD-DBM peptide promotes formation of neogenic hair follicles (Figure 8B), proved by the presence of white hair in the healed tissues.¹²⁴ Recent clinical studies by the Clinical Peptide Society found that PTD-DBM peptide enhances human hair growth. These observations indicate that activation of the Wnt/ β -catenin signaling by the blockade of the CXXC5–DVL interaction with PTD-DBM peptide could be a therapeutic strategy for regenerative wound healing.

Perspectives

The Wnt/ β -catenin pathway is an attractive target for regenerative wound healing. Although agents activating Wnt/ β -catenin signaling stimulate the wound healing process, an appropriate dosing and treatment duration is important because aberrant activation of the Wnt/ β -catenin pathway causes fibrotic diseases including hypertrophic scarring, keloid formation, and skin cancer. However, activating Wnt/ β -catenin signaling via releasing the CXXC5-mediated negative feedback loop instead of via direct activation enhances wound healing without leading to unwanted outcomes such as melanoma. The safety of this therapeutic approach was confirmed by the absence of any pathological skin phenotypes, including melanoma-accompanying transformations, in 1-year-old *Cxhc5*^{-/-} mice or in mice that received topical application of PTD-DBM peptide for more than 6 months. Furthermore, PTD-DBM peptide treatment does not induce transcription of *cyclin D1* and *c-Myc*, which are the Wnt/ β -catenin signaling target genes frequently overexpressed during cancer development, but does induce transcription of *endothelin-1*, which contributes to enhanced wound healing.¹³ Finally, the specific blockade of the cytosolic function suppressing Wnt/ β -catenin signaling, not the nuclear function acting as a transcription factor,^{119,120} further provides conceptual safety of an approach interfering the CXXC5–DVL interaction for target specificity. Therefore, the CXXC5–DVL interaction is potentially a safe target for regenerative wound healing. A strategy to discover small molecules mimicking the PTD-DBM

peptide could be a valuable approach for development of a first-in-class wound healing agents that would be cost-effective and suitable for routine use.

6.0 SUMMARY

An acute skin wound is spontaneously repaired within 1–2 weeks. However, due to scar formation, the repaired skin is not identical to intact uninjured skin. As the elderly population with delayed wound healing increases, the need for effective wound healing agents based on regenerative healing also increases. Current wound care research has focused on regenerative therapies to diminish scar formation, improve the quality of restored skin, and accelerate healing rates.

The Wnt/ β -catenin pathway could be as a major target for development of drugs in the field of regenerative wound healing because it can promote the overall wound healing process by activation of stem cells through interaction with other signaling pathways including the TGF β /Smad pathway.

Activators of Wnt/ β -catenin signaling, such as small molecules (e.g. LiCl and VPA) and natural products (e.g. lucidone and *Polygonum aviculare L.* extract), have been characterized as agents enhancing wound healing. However, their effectiveness during the healing process could be restricted due to induction of CXXC5, a negative feedback regulator of the Wnt/ β -catenin pathway, during the early stages of wound healing. Therefore, inhibition of CXXC5 function, especially the cytosolic form that suppresses Wnt/ β -catenin signaling via interaction with DVL (CXXC5–DVL interaction, PPI), is a target for the development of novel regenerative wound healing agents. Topically applied PTD-DBM peptide, which interferes with the CXXC5–DVL interaction, effectively enhances the wound healing process and has potential as a therapeutic agent. The maximal effects of PTD-DBM peptide occur in combination with direct Wnt/ β -catenin signaling activators, such as VPA. This combination treatment promotes regenerative wound healing via strong activation of the Wnt/ β -catenin pathway; initial activation occurs through VPA-induced GSK-3 β inactivation. Subsequent further activation is enhanced via blockade of the CXXC5–DVL interaction by the PTD-DBM peptide. This approach for wound healing activating the Wnt/ β -catenin signaling could minimize undesirable side effects (e.g., skin cancer) that can be induced via aberrant activation of the Wnt/ β -catenin pathway.

TAKE-HOME MESSAGES

- Use of regenerative therapies including stem cells and growth factors is a current approach for complete healing of skin wounds.
- Multiple signaling pathways are involved in the skin wound healing, and the Wnt/ β -catenin pathway is a key player in the wound healing process.
- The Wnt/ β -catenin pathway participates in the activation of skin stem cells as well as the overall process of wound healing to enhance regenerative wound healing.
- CXXC5, a negative feedback regulator of the Wnt/ β -catenin pathway, suppresses wound healing by exerting its function via suppression of this pathway.
- Inhibition of CXXC5 function via its binding to DVL enhances the speed and quality of healing in mouse skin wounds, without any adverse effects.
- Restoration of the Wnt/ β -catenin signaling via blockade of the CXXC5-mediated negative feedback mechanism, not by direct activation, is a potential future strategy for regenerative wound healing.

ACKNOWLEDGMENTS AND FUNDING SOURCES

This work was supported by the National Research Foundation (NRF) of Korea grant funded by the Korean Government (MSIP) (2019R1A2C3002751, 2020M3E5E2040018). M.Y. was supported by a BK21 PLUS program.

AUTHOR CONTRIBUTIONS

K.C. conceptualized and supervised the project. S.C. and M.Y. wrote the manuscript and drew the figures. S.C. and K.C. revised the manuscript.

AUTHOR DISCLOSURE AND GHOSTWRITING

No competing financial interests exist. The content of this article was entirely written by the authors listed. No ghostwriters were used to write this article.

ABOUT THE AUTHORS

Sehee Choi, PhD, received a PhD degree from Yonsei University.

Minguen Yoon is a PhD student in the Department of Biotechnology at Yonsei University.

Kang-Yell Choi, PhD, is a Professor in the Department of Biotechnology at Yonsei University. He also serves as the CEO of CK Biotech Inc., headquartered in Seoul, Korea, which has a license to develop and use the compounds and the peptide disclosed in this article.

ABBREVIATIONS AND ACRONYMS

ALP = alkaline phosphatase

APC = adenomatous polyposis coli

CK1 = casein kinase 1

CXXC5 = CXXC-type zinc finger protein 5

DBM = dishevelled binding motif

DVL =dishevelled

ECM = extracellular matrix

EGF = epidermal growth factor

ESC = epidermal stem cell

EpSC = epithelial stem cell

FDA = Food and Drug Administration

FGF = fibroblast growth factor

GSK-3 = glycogen synthase kinase-3

HA = hyaluronic acid

HSC= hematopoietic stem cell

IL = interleukin

iPSC = induced pluripotent stem cell

LGR5 = leucine rich repeat containing G protein-coupled receptor 5

LiCl = lithium chloride

MMP = metalloproteinases

MSC = mesenchymal stem cell

PDGF = platelet-derived growth factor

PPI = protein–protein interaction

PTD =protein transduction domain

TGF- β = transforming growth factor beta

TNF = tumor necrosis factor

VEGF = vascular endothelial growth factor

VPA = valproic acid

WISP1 = Wnt1-inducible signaling pathway protein 1

REFERENCES

1. Reinke JM, Sorg H. Wound repair and regeneration. *Eur Surg Res.* 2012;49(1):35-43.
2. Velnar T, Bailey T, Smrkolj V. The wound healing process: an overview of the cellular and molecular mechanisms. *J Int Med Res.* Sep-Oct 2009;37(5):1528-1542.
3. Sen CK, Gordillo GM, Roy S, et al. Human skin wounds: a major and snowballing threat to public health and the economy. *Wound Repair Regen.* Nov-Dec 2009;17(6):763-771.
4. Shah JB. The history of wound care. *J Am Col Certif Wound Spec.* Sep 2011;3(3):65-66.
5. Takeo M, Lee W, Ito M. Wound healing and skin regeneration. *Cold Spring Harb Perspect Med.* Jan 5 2015;5(1):a023267.
6. Gurtner GC, Werner S, Barrandon Y, Longaker MT. Wound repair and regeneration. *Nature.* May 15 2008;453(7193):314-321.
7. Marshall CD, Hu MS, Leavitt T, Barnes LA, Lorenz HP, Longaker MT. Cutaneous Scarring: Basic Science, Current Treatments, and Future Directions. *Adv Wound Care (New Rochelle).* Feb 1 2018;7(2):29-45.
8. Werner S, Grose R. Regulation of wound healing by growth factors and cytokines. *Physiol Rev.* Jul 2003;83(3):835-870.
9. Lau K, Paus R, Tiede S, Day P, Bayat A. Exploring the role of stem cells in cutaneous wound healing. *Exp Dermatol.* Nov 2009;18(11):921-933.
10. Volk SW, Theoret C. Translating stem cell therapies: the role of companion animals in regenerative medicine. *Wound Repair Regen.* May-Jun 2013;21(3):382-394.
11. Whittam AJ, Maan ZN, Duscher D, et al. Challenges and Opportunities in Drug Delivery for Wound Healing. *Adv Wound Care (New Rochelle).* Feb 1 2016;5(2):79-88.
12. Bielefeld KA, Amini-Nik S, Alman BA. Cutaneous wound healing: recruiting developmental pathways for regeneration. *Cell Mol Life Sci.* Jun 2013;70(12):2059-2081.
13. Lee SH, Kim MY, Kim HY, et al. The Dishevelled-binding protein CXXC5 negatively regulates cutaneous wound healing. *J Exp Med.* Jun 29 2015;212(7):1061-1080.

14. Li J, Chen J, Kirsner R. Pathophysiology of acute wound healing. *Clin Dermatol*. Jan-Feb 2007;25(1):9-18.
15. Mack M. Inflammation and fibrosis. *Matrix Biol*. Aug 2018;68-69:106-121.
16. Xu X, Gu S, Huang X, et al. The role of macrophages in the formation of hypertrophic scars and keloids. *Burns Trauma*. 2020;8:tkaa006.
17. Rowlatt U. Intrauterine wound healing in a 20 week human fetus. *Virchows Arch A Pathol Anat Histol*. Mar 23 1979;381(3):353-361.
18. Whitby DJ, Ferguson MW. The extracellular matrix of lip wounds in fetal, neonatal and adult mice. *Development*. Jun 1991;112(2):651-668.
19. Ferguson MW, O'Kane S. Scar-free healing: from embryonic mechanisms to adult therapeutic intervention. *Philos Trans R Soc Lond B Biol Sci*. May 29 2004;359(1445):839-850.
20. Londono R, Sun AX, Tuan RS, Lozito TP. Tissue Repair and Epimorphic Regeneration: An Overview. *Curr Pathobiol Rep*. Mar 2018;6(1):61-69.
21. Iismaa SE, Kaidonis X, Nicks AM, et al. Comparative regenerative mechanisms across different mammalian tissues. *NPJ Regen Med*. 2018;3:6.
22. Ito M, Yang Z, Andl T, et al. Wnt-dependent de novo hair follicle regeneration in adult mouse skin after wounding. *Nature*. May 17 2007;447(7142):316-320.
23. Whyte JL, Smith AA, Helms JA. Wnt signaling and injury repair. *Cold Spring Harb Perspect Biol*. Aug 1 2012;4(8):a008078.
24. Houschyar KS, Momeni A, Pyles MN, Maan ZN, Whittam AJ, Siemers F. Wnt signaling induces epithelial differentiation during cutaneous wound healing. *Organogenesis*. 2015;11(3):95-104.
25. Logan CY, Nusse R. The Wnt signaling pathway in development and disease. *Annu Rev Cell Dev Biol*. 2004;20:781-810.
26. Reya T, Clevers H. Wnt signalling in stem cells and cancer. *Nature*. Apr 14 2005;434(7035):843-850.
27. Pierce GF, Mustoe TA, Altrock BW, Deuel TF, Thomason A. Role of platelet-derived growth factor in wound healing. *J Cell Biochem*. Apr 1991;45(4):319-326.

28. Lynch SE, Nixon JC, Colvin RB, Antoniades HN. Role of platelet-derived growth factor in wound healing: synergistic effects with other growth factors. *Proc Natl Acad Sci U S A*. Nov 1987;84(21):7696-7700.
29. LeGrand EK. Preclinical promise of becaplermin (rhPDGF-BB) in wound healing. *Am J Surg*. Aug 1998;176(2A Suppl):48S-54S.
30. Chan RK, Liu PH, Pietramaggiore G, Ibrahim SI, Hechtman HB, Orgill DP. Effect of recombinant platelet-derived growth factor (Regranex) on wound closure in genetically diabetic mice. *J Burn Care Res*. Mar-Apr 2006;27(2):202-205.
31. Park SA, Raghunathan VK, Shah NM, et al. PDGF-BB does not accelerate healing in diabetic mice with splinted skin wounds. *PLoS One*. 2014;9(8):e104447.
32. Barrandon Y, Green H. Cell migration is essential for sustained growth of keratinocyte colonies: the roles of transforming growth factor-alpha and epidermal growth factor. *Cell*. Sep 25 1987;50(7):1131-1137.
33. Marikovsky M, Breuing K, Liu PY, et al. Appearance of heparin-binding EGF-like growth factor in wound fluid as a response to injury. *Proc Natl Acad Sci U S A*. May 1 1993;90(9):3889-3893.
34. Brown GL, Nanney LB, Griffen J, et al. Enhancement of wound healing by topical treatment with epidermal growth factor. *N Engl J Med*. Jul 13 1989;321(2):76-79.
35. Khanbanha N, Atyabi F, Taheri A, Talaie F, Mahbod M, Dinarvand R. Healing efficacy of an EGF impregnated triple gel based wound dressing: in vitro and in vivo studies. *Biomed Res Int*. 2014;2014:493732.
36. Lee JH, Bae IH, Choi JK, Park JW. Evaluation of a highly skin permeable low-molecular-weight protamine conjugated epidermal growth factor for novel burn wound healing therapy. *J Pharm Sci*. Nov 2013;102(11):4109-4120.
37. Amendt C, Mann A, Schirmacher P, Blessing M. Resistance of keratinocytes to TGFbeta-mediated growth restriction and apoptosis induction accelerates re-epithelialization in skin wounds. *J Cell Sci*. May 15 2002;115(Pt 10):2189-2198.
38. Finnsen KW, Arany PR, Philip A. Transforming Growth Factor Beta Signaling in Cutaneous Wound Healing: Lessons Learned from Animal Studies. *Adv Wound Care (New Rochelle)*. Jun 2013;2(5):225-237.

39. McCollum PT, Bush JA, James G, et al. Randomized phase II clinical trial of avotermin versus placebo for scar improvement. *Br J Surg*. Jul 2011;98(7):925-934.
40. So K, McGrouther DA, Bush JA, et al. Avotermin for scar improvement following scar revision surgery: a randomized, double-blind, within-patient, placebo-controlled, phase II clinical trial. *Plast Reconstr Surg*. Jul 2011;128(1):163-172.
41. Zhou K, Ma Y, Brogan MS. Chronic and non-healing wounds: The story of vascular endothelial growth factor. *Med Hypotheses*. Oct 2015;85(4):399-404.
42. Eming SA, Krieg T. Molecular mechanisms of VEGF-A action during tissue repair. *J Invest Dermatol Symp Proc*. Sep 2006;11(1):79-86.
43. Hanft JR, Pollak RA, Barbul A, et al. Phase I trial on the safety of topical rhVEGF on chronic neuropathic diabetic foot ulcers. *J Wound Care*. Jan 2008;17(1):30-32, 34-37.
44. Shi HX, Lin C, Lin BB, et al. The anti-scar effects of basic fibroblast growth factor on the wound repair in vitro and in vivo. *PLoS One*. 2013;8(4):e59966.
45. Nakamizo S, Egawa G, Doi H, Natsuaki Y, Miyachi Y, Kabashima K. Topical treatment with basic fibroblast growth factor promotes wound healing and barrier recovery induced by skin abrasion. *Skin Pharmacol Physiol*. 2013;26(1):22-29.
46. Robson MC, Phillips TJ, Falanga V, et al. Randomized trial of topically applied repifermin (recombinant human keratinocyte growth factor-2) to accelerate wound healing in venous ulcers. *Wound Repair Regen*. Sep-Oct 2001;9(5):347-352.
47. Ma B, Cheng DS, Xia ZF, et al. Randomized, multicenter, double-blind, and placebo-controlled trial using topical recombinant human acidic fibroblast growth factor for deep partial-thickness burns and skin graft donor site. *Wound Repair Regen*. Nov-Dec 2007;15(6):795-799.
48. Andree C, Swain WF, Page CP, et al. In vivo transfer and expression of a human epidermal growth factor gene accelerates wound repair. *Proc Natl Acad Sci U S A*. Dec 6 1994;91(25):12188-12192.
49. Bodnar RJ. Epidermal Growth Factor and Epidermal Growth Factor Receptor: The Yin and Yang in the Treatment of Cutaneous Wounds and Cancer. *Adv Wound Care (New Rochelle)*. Feb 2013;2(1):24-29.

50. Kim YS, Sung DK, Kong WH, Kim H, Hahn SK. Synergistic effects of hyaluronate - epidermal growth factor conjugate patch on chronic wound healing. *Biomater Sci*. May 1 2018;6(5):1020-1030.
51. Frank S, Madlener M, Werner S. Transforming growth factors beta1, beta2, and beta3 and their receptors are differentially regulated during normal and impaired wound healing. *J Biol Chem*. Apr 26 1996;271(17):10188-10193.
52. Fu X, Li H. Mesenchymal stem cells and skin wound repair and regeneration: possibilities and questions. *Cell Tissue Res*. Feb 2009;335(2):317-321.
53. Wu Y, Chen L, Scott PG, Tredget EE. Mesenchymal stem cells enhance wound healing through differentiation and angiogenesis. *Stem Cells*. Oct 2007;25(10):2648-2659.
54. Chen L, Tredget EE, Wu PY, Wu Y. Paracrine factors of mesenchymal stem cells recruit macrophages and endothelial lineage cells and enhance wound healing. *PLoS One*. Apr 2 2008;3(4):e1886.
55. Feldman DS, McCauley JF. Mesenchymal Stem Cells and Transforming Growth Factor-beta(3) (TGF-beta(3)) to Enhance the Regenerative Ability of an Albumin Scaffold in Full Thickness Wound Healing. *J Funct Biomater*. Nov 14 2018;9(4).
56. Nauta AJ, Fibbe WE. Immunomodulatory properties of mesenchymal stromal cells. *Blood*. Nov 15 2007;110(10):3499-3506.
57. Isakson M, de Blacam C, Whelan D, McArdle A, Clover AJ. Mesenchymal Stem Cells and Cutaneous Wound Healing: Current Evidence and Future Potential. *Stem Cells Int*. 2015;2015:831095.
58. Krause DS, Theise ND, Collector MI, et al. Multi-organ, multi-lineage engraftment by a single bone marrow-derived stem cell. *Cell*. May 4 2001;105(3):369-377.
59. Chan RK, Garfein E, Gigante PR, et al. Side population hematopoietic stem cells promote wound healing in diabetic mice. *Plast Reconstr Surg*. Aug 2007;120(2):407-411; discussion 412-403.
60. Plikus MV, Gay DL, Treffeisen E, Wang A, Supapannachart RJ, Cotsarelis G. Epithelial stem cells and implications for wound repair. *Semin Cell Dev Biol*. Dec 2012;23(9):946-953.

61. Yang X, Qu L, Wang X, et al. Plasticity of epidermal adult stem cells derived from adult goat ear skin. *Mol Reprod Dev.* Mar 2007;74(3):386-396.
62. Broeckx SY, Maes S, Martinello T, et al. Equine epidermis: a source of epithelial-like stem/progenitor cells with in vitro and in vivo regenerative capacities. *Stem Cells Dev.* May 15 2014;23(10):1134-1148.
63. Doulatov S, Daley GQ. Development. A stem cell perspective on cellular engineering. *Science.* Nov 8 2013;342(6159):700-702.
64. Cao Y, Gang X, Sun C, Wang G. Mesenchymal Stem Cells Improve Healing of Diabetic Foot Ulcer. *J Diabetes Res.* 2017;2017:9328347.
65. Ferraro F, Celso CL, Scadden D. Adult stem cells and their niches. *Adv Exp Med Biol.* 2010;695:155-168.
66. Tanabe S. Signaling involved in stem cell reprogramming and differentiation. *World J Stem Cells.* Aug 26 2015;7(7):992-998.
67. Soo C, Beanes SR, Hu FY, et al. Ontogenetic transition in fetal wound transforming growth factor-beta regulation correlates with collagen organization. *Am J Pathol.* Dec 2003;163(6):2459-2476.
68. Shah M, Foreman DM, Ferguson MW. Neutralisation of TGF-beta 1 and TGF-beta 2 or exogenous addition of TGF-beta 3 to cutaneous rat wounds reduces scarring. *J Cell Sci.* Mar 1995;108 (Pt 3):985-1002.
69. Owens P, Han G, Li AG, Wang XJ. The role of Smads in skin development. *J Invest Dermatol.* Apr 2008;128(4):783-790.
70. Oshimori N, Fuchs E. Paracrine TGF-beta signaling counterbalances BMP-mediated repression in hair follicle stem cell activation. *Cell Stem Cell.* Jan 6 2012;10(1):63-75.
71. Blanpain C, Fuchs E. Epidermal homeostasis: a balancing act of stem cells in the skin. *Nat Rev Mol Cell Biol.* Mar 2009;10(3):207-217.
72. Okuyama R, Tagami H, Aiba S. Notch signaling: its role in epidermal homeostasis and in the pathogenesis of skin diseases. *J Dermatol Sci.* Mar 2008;49(3):187-194.
73. Watt FM, Estrach S, Ambler CA. Epidermal Notch signalling: differentiation, cancer and adhesion. *Curr Opin Cell Biol.* Apr 2008;20(2):171-179.
74. Gridley T. Notch signaling in the vasculature. *Curr Top Dev Biol.* 2010;92:277-309.

75. Asai J, Takenaka H, Kusano KF, et al. Topical sonic hedgehog gene therapy accelerates wound healing in diabetes by enhancing endothelial progenitor cell-mediated microvascular remodeling. *Circulation*. May 23 2006;113(20):2413-2424.
76. Le H, Kleinerman R, Lerman OZ, et al. Hedgehog signaling is essential for normal wound healing. *Wound Repair Regen*. Nov-Dec 2008;16(6):768-773.
77. Clevers H, Loh KM, Nusse R. Stem cell signaling. An integral program for tissue renewal and regeneration: Wnt signaling and stem cell control. *Science*. Oct 3 2014;346(6205):1248012.
78. Bastakoty D, Young PP. Wnt/beta-catenin pathway in tissue injury: roles in pathology and therapeutic opportunities for regeneration. *FASEB J*. Oct 2016;30(10):3271-3284.
79. Steinhart Z, Angers S. Wnt signaling in development and tissue homeostasis. *Development*. Jun 8 2018;145(11).
80. Kim HY, Yoon JY, Yun JH, et al. CXXC5 is a negative-feedback regulator of the Wnt/beta-catenin pathway involved in osteoblast differentiation. *Cell Death Differ*. Jun 2015;22(6):912-920.
81. Miki T, Yasuda SY, Kahn M. Wnt/beta-catenin signaling in embryonic stem cell self-renewal and somatic cell reprogramming. *Stem Cell Rev*. Nov 2011;7(4):836-846.
82. Kawakami Y, Rodriguez Esteban C, Raya M, et al. Wnt/beta-catenin signaling regulates vertebrate limb regeneration. *Genes Dev*. Dec 1 2006;20(23):3232-3237.
83. Nusse R, Clevers H. Wnt/beta-Catenin Signaling, Disease, and Emerging Therapeutic Modalities. *Cell*. Jun 1 2017;169(6):985-999.
84. MacDonald BT, Tamai K, He X. Wnt/beta-catenin signaling: components, mechanisms, and diseases. *Dev Cell*. Jul 2009;17(1):9-26.
85. Moon RT, Kohn AD, De Ferrari GV, Kaykas A. WNT and beta-catenin signalling: diseases and therapies. *Nat Rev Genet*. Sep 2004;5(9):691-701.
86. Mazumdar J, O'Brien WT, Johnson RS, et al. O2 regulates stem cells through Wnt/beta-catenin signalling. *Nat Cell Biol*. Oct 2010;12(10):1007-1013.
87. Poon R, Nik SA, Ahn J, Slade L, Alman BA. Beta-catenin and transforming growth factor beta have distinct roles regulating fibroblast cell motility and the induction of collagen lattice contraction. *BMC Cell Biol*. May 11 2009;10:38.

88. Cheon S, Poon R, Yu C, et al. Prolonged beta-catenin stabilization and tcf-dependent transcriptional activation in hyperplastic cutaneous wounds. *Lab Invest*. Mar 2005;85(3):416-425.
89. Sun Z, Wang C, Shi C, et al. Activated Wnt signaling induces myofibroblast differentiation of mesenchymal stem cells, contributing to pulmonary fibrosis. *Int J Mol Med*. May 2014;33(5):1097-1109.
90. Birdsey GM, Shah AV, Dufton N, et al. The endothelial transcription factor ERG promotes vascular stability and growth through Wnt/beta-catenin signaling. *Dev Cell*. Jan 12 2015;32(1):82-96.
91. Lim X, Tan SH, Koh WL, et al. Interfollicular epidermal stem cells self-renew via autocrine Wnt signaling. *Science*. Dec 6 2013;342(6163):1226-1230.
92. Merrill BJ, Gat U, DasGupta R, Fuchs E. Tcf3 and Lef1 regulate lineage differentiation of multipotent stem cells in skin. *Genes Dev*. Jul 1 2001;15(13):1688-1705.
93. Veltri A, Lang C, Lien WH. Concise Review: Wnt Signaling Pathways in Skin Development and Epidermal Stem Cells. *Stem Cells*. Jan 2018;36(1):22-35.
94. Shi Y, Shu B, Yang R, et al. Wnt and Notch signaling pathway involved in wound healing by targeting c-Myc and Hes1 separately. *Stem Cell Res Ther*. Jun 16 2015;6:120.
95. Fuchs E. Scratching the surface of skin development. *Nature*. Feb 22 2007;445(7130):834-842.
96. Silva-Vargas V, Lo Celso C, Giangreco A, et al. Beta-catenin and Hedgehog signal strength can specify number and location of hair follicles in adult epidermis without recruitment of bulge stem cells. *Dev Cell*. Jul 2005;9(1):121-131.
97. Chuong CM. Regenerative biology: new hair from healing wounds. *Nature*. May 17 2007;447(7142):265-266.
98. Smith AA, Li J, Liu B, et al. Activating Hair Follicle Stem Cells via R-spondin2 to Stimulate Hair Growth. *J Invest Dermatol*. Aug 2016;136(8):1549-1558.
99. Buchanan EP, Longaker MT, Lorenz HP. Fetal skin wound healing. *Adv Clin Chem*. 2009;48:137-161.

100. West DC, Shaw DM, Lorenz P, Adzick NS, Longaker MT. Fibrotic healing of adult and late gestation fetal wounds correlates with increased hyaluronidase activity and removal of hyaluronan. *Int J Biochem Cell Biol.* Jan 1997;29(1):201-210.
101. Sobel K, Tham M, Stark HJ, et al. Wnt-3a-activated human fibroblasts promote human keratinocyte proliferation and matrix destruction. *Int J Cancer.* Jun 15 2015;136(12):2786-2798.
102. Jordan AR, Racine RR, Hennig MJ, Lokeshwar VB. The Role of CD44 in Disease Pathophysiology and Targeted Treatment. *Front Immunol.* 2015;6:182.
103. Savani RC, Cao G, Pooler PM, Zaman A, Zhou Z, DeLisser HM. Differential involvement of the hyaluronan (HA) receptors CD44 and receptor for HA-mediated motility in endothelial cell function and angiogenesis. *J Biol Chem.* Sep 28 2001;276(39):36770-36778.
104. Xue M, Jackson CJ. Extracellular Matrix Reorganization During Wound Healing and Its Impact on Abnormal Scarring. *Adv Wound Care (New Rochelle).* Mar 1 2015;4(3):119-136.
105. Akhmetshina A, Palumbo K, Dees C, et al. Activation of canonical Wnt signalling is required for TGF-beta-mediated fibrosis. *Nat Commun.* Mar 13 2012;3:735.
106. Carthy JM, Garmaroudi FS, Luo Z, McManus BM. Wnt3a induces myofibroblast differentiation by upregulating TGF-beta signaling through SMAD2 in a beta-catenin-dependent manner. *PLoS One.* 2011;6(5):e19809.
107. Liu J, Wang Y, Pan Q, et al. Wnt/beta-catenin pathway forms a negative feedback loop during TGF-beta1 induced human normal skin fibroblast-to-myofibroblast transition. *J Dermatol Sci.* Jan 2012;65(1):38-49.
108. Gay D, Kwon O, Zhang Z, et al. Fgf9 from dermal gammadelta T cells induces hair follicle neogenesis after wounding. *Nat Med.* Jul 2013;19(7):916-923.
109. Huang P, Yan R, Zhang X, Wang L, Ke X, Qu Y. Activating Wnt/beta-catenin signaling pathway for disease therapy: Challenges and opportunities. *Pharmacol Ther.* Apr 2019;196:79-90.
110. Tran FH, Zheng JJ. Modulating the wnt signaling pathway with small molecules. *Protein Sci.* Apr 2017;26(4):650-661.

111. Kramer T, Schmidt B, Lo Monte F. Small-Molecule Inhibitors of GSK-3: Structural Insights and Their Application to Alzheimer's Disease Models. *Int J Alzheimers Dis.* 2012;2012:381029.
112. Lee SH, Zahoor M, Hwang JK, Min do S, Choi KY. Valproic acid induces cutaneous wound healing in vivo and enhances keratinocyte motility. *PLoS One.* 2012;7(11):e48791.
113. Yang HL, Tsai YC, Korivi M, Chang CT, Hseu YC. Lucidone Promotes the Cutaneous Wound Healing Process via Activation of the PI3K/AKT, Wnt/beta-catenin and NF-kappaB Signaling Pathways. *Biochim Biophys Acta Mol Cell Res.* Jan 2017;1864(1):151-168.
114. Cha PH, Shin W, Zahoor M, Kim HY, Min do S, Choi KY. Hovenia dulcis Thunb extract and its ingredient methyl vanillate activate Wnt/beta-catenin pathway and increase bone mass in growing or ovariectomized mice. *PLoS One.* 2014;9(1):e85546.
115. Seo SH, Lee SH, Cha PH, Kim MY, Min do S, Choi KY. Polygonum aviculare L. and its active compounds, quercitrin hydrate, caffeic acid, and rutin, activate the Wnt/beta-catenin pathway and induce cutaneous wound healing. *Phytother Res.* May 2016;30(5):848-854.
116. Kabashima K, Sakabe J, Yoshiki R, Tabata Y, Kohno K, Tokura Y. Involvement of Wnt signaling in dermal fibroblasts. *Am J Pathol.* Feb 2010;176(2):721-732.
117. Chua AW, Gan SU, Ting Y, et al. Keloid fibroblasts are more sensitive to Wnt3a treatment in terms of elevated cellular growth and fibronectin expression. *J Dermatol Sci.* Dec 2011;64(3):199-209.
118. Li CH, Amar S. Role of secreted frizzled-related protein 1 (SFRP1) in wound healing. *J Dent Res.* Apr 2006;85(4):374-378.
119. Kim HY, Yang DH, Shin SW, et al. CXXC5 is a transcriptional activator of Flk-1 and mediates bone morphogenic protein-induced endothelial cell differentiation and vessel formation. *FASEB J.* Feb 2014;28(2):615-626.
120. Kim MY, Kim HY, Hong J, et al. CXXC5 plays a role as a transcription activator for myelin genes on oligodendrocyte differentiation. *Glia.* Mar 2016;64(3):350-362.

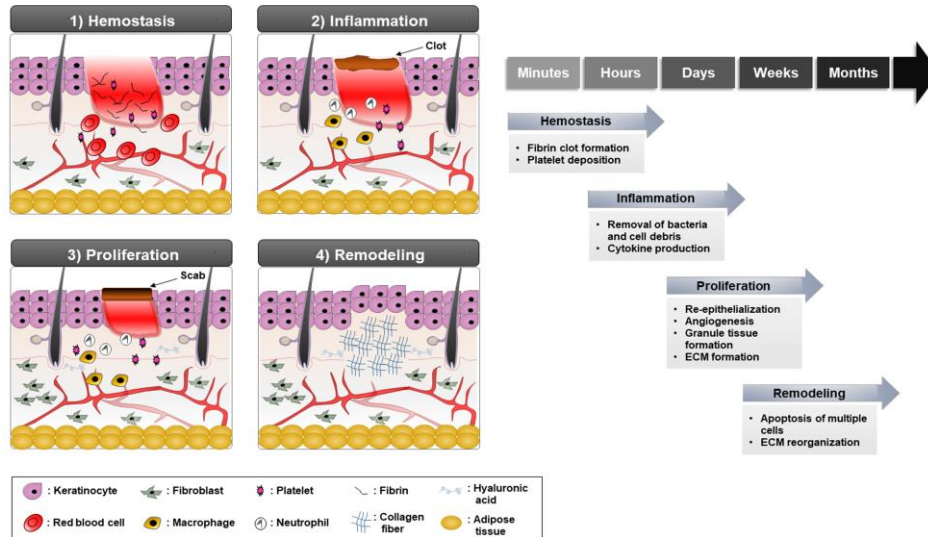
121. Kim MS, Yoon SK, Bollig F, et al. A novel Wilms tumor 1 (WT1) target gene negatively regulates the WNT signaling pathway. *J Biol Chem*. May 7 2010;285(19):14585-14593.
122. Gao C, Chen YG. Dishevelled: The hub of Wnt signaling. *Cell Signal*. May 2010;22(5):717-727.
123. Andersson T, Sodersten E, Duckworth JK, et al. CXXC5 is a novel BMP4-regulated modulator of Wnt signaling in neural stem cells. *J Biol Chem*. Feb 6 2009;284(6):3672-3681.
124. Lee SH, Seo SH, Lee DH, Pi LQ, Lee WS, Choi KY. Targeting of CXXC5 by a Competing Peptide Stimulates Hair Regrowth and Wound-Induced Hair Neogenesis. *J Invest Dermatol*. Nov 2017;137(11):2260-2269.
125. Choi S, Kim HY, Cha PH, et al. CXXC5 mediates growth plate senescence and is a target for enhancement of longitudinal bone growth. *Life Sci Alliance*. Apr 2019;2(2).
126. Lim X, Tan SH, Yu KL, Lim SB, Nusse R. Axin2 marks quiescent hair follicle bulge stem cells that are maintained by autocrine Wnt/beta-catenin signaling. *Proc Natl Acad Sci U S A*. Mar 15 2016;113(11):E1498-1505.
127. Lim CH, Sun Q, Ratti K, et al. Hedgehog stimulates hair follicle neogenesis by creating inductive dermis during murine skin wound healing. *Nat Commun*. Nov 21 2018;9(1):4903.
128. Haukipuro K, Melkko J, Risteli L, Kairaluoma M, Risteli J. Synthesis of type I collagen in healing wounds in humans. *Ann Surg*. Jan 1991;213(1):75-80.
129. Shaw TJ, Martin P. Wound repair at a glance. *J Cell Sci*. Sep 15 2009;122(Pt 18):3209-3213.
130. Volk SW, Wang Y, Mauldin EA, Liechty KW, Adams SL. Diminished type III collagen promotes myofibroblast differentiation and increases scar deposition in cutaneous wound healing. *Cells Tissues Organs*. 2011;194(1):25-37.
131. Repertinger SK, Campagnaro E, Fuhrman J, El-Abaseri T, Yuspa SH, Hansen LA. EGFR enhances early healing after cutaneous incisional wounding. *J Invest Dermatol*. Nov 2004;123(5):982-989.

132. Lagares D, Garcia-Fernandez RA, Jimenez CL, et al. Endothelin 1 contributes to the effect of transforming growth factor beta1 on wound repair and skin fibrosis. *Arthritis Rheum.* Mar 2010;62(3):878-889.
133. Lenselink EA. Role of fibronectin in normal wound healing. *Int Wound J.* Jun 2015;12(3):313-316.
134. Grinnell F. Fibronectin and wound healing. *J Cell Biochem.* 1984;26(2):107-116.
135. Chan T, Ghahary A, Demare J, et al. Development, characterization, and wound healing of the keratin 14 promoted transforming growth factor-beta1 transgenic mouse. *Wound Repair Regen.* May-Jun 2002;10(3):177-187.
136. Bao P, Kodra A, Tomic-Canic M, Golinko MS, Ehrlich HP, Brem H. The role of vascular endothelial growth factor in wound healing. *J Surg Res.* May 15 2009;153(2):347-358.
137. Johnson KE, Wilgus TA. Vascular Endothelial Growth Factor and Angiogenesis in the Regulation of Cutaneous Wound Repair. *Adv Wound Care (New Rochelle).* Oct 1 2014;3(10):647-661.
138. Lu S, Liu H, Lu L, et al. WISP1 overexpression promotes proliferation and migration of human vascular smooth muscle cells via AKT signaling pathway. *Eur J Pharmacol.* Oct 5 2016;788:90-97.
139. Ono M, Masaki A, Maeda A, et al. CCN4/WISP1 controls cutaneous wound healing by modulating proliferation, migration and ECM expression in dermal fibroblasts via alpha5beta1 and TNFalpha. *Matrix Biol.* Aug 2018;68-69:533-546.
140. Qi W, Yang C, Dai Z, et al. High levels of pigment epithelium-derived factor in diabetes impair wound healing through suppression of Wnt signaling. *Diabetes.* Apr 2015;64(4):1407-1419.
141. Driskell RR, Lichtenberger BM, Hoste E, et al. Distinct fibroblast lineages determine dermal architecture in skin development and repair. *Nature.* Dec 12 2013;504(7479):277-281.
142. Popp T, Steinritz D, Breit A, et al. Wnt5a/beta-catenin signaling drives calcium-induced differentiation of human primary keratinocytes. *J Invest Dermatol.* Aug 2014;134(8):2183-2191.

143. Zhu XJ, Liu Y, Dai ZM, et al. BMP-FGF signaling axis mediates Wnt-induced epidermal stratification in developing mammalian skin. *PLoS Genet.* Oct 2014;10(10):e1004687.
144. Bukowska J, Walendzik K, Kopcewicz M, Cierniak P, Gawronska-Kozak B. Wnt signaling and the transcription factor Foxn1 contribute to cutaneous wound repair in mice. *Connect Tissue Res.* Nov 10 2019:1-11.

FIGURE LEGENDS

Figure 1

**Figure 1. Wound repair phases**

The four different stages of the wound repair process. **1)** The hemostasis phase (begins immediately after wounding), the coagulation process blocks the current leakage of blood and fluids via fibrin network formation and platelet deposition. This initial phase also has a role as a barricade against microorganism entry into the lesion. **2)** The inflammatory phase (begins within 24 hours after wound formation and lasts for several days), neutrophils and macrophages remove bacteria and cell debris. They also promote the production of cytokines and assist other inflammatory cells recruited to the wound region. **3)** The proliferative phase (begins 4–5 days after wound formation and lasts for several weeks), re-epithelialization, angiogenesis, and extracellular matrix (ECM) and granulation tissue formation occur via the activation of, and crosstalk between, multiple signaling cascades. **4)** The final remodeling phase (begins at approximately 3 weeks after wound formation and lasts for as long as 1–2 years), the tissue tensile strength is generated by ECM reorganization. Multiple cells undergo apoptosis to finish tissue remodeling.

Figure 2

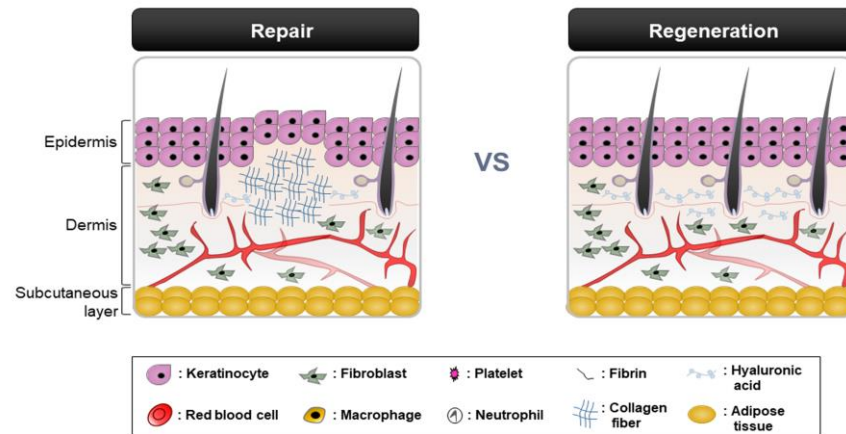


Figure 2. Comparison between repair and regeneration in skin wound healing

Left, Repaired skin. Healing by the repair process fails to restore skin to uninjured status and remains scar due to the alignment of excessive collagen fibers in the dermis. Right, Regenerated skin. The regenerative healing induces *de novo* synthesis of hyaluronic acid, and results in complete restoration of skin tissue with the formation of adnexa including hair.

Figure 3

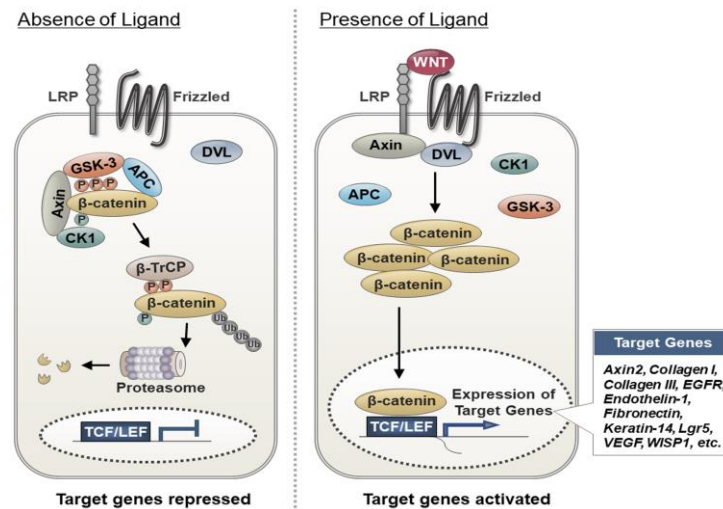


Figure 3. Wnt/β-catenin pathway and its target genes

In absence of the Wnt ligand, the destruction complex composed of Axin, APC (adenomatous polyposis coli), GSK-3 (glycogen synthase kinase-3), and CK1 (casein kinase 1) is formed in the cytoplasm. By forming this complex, β-catenin is phosphorylated initially by CK1 and subsequently by GSK-3. The β-TrCP E3 linker is recruited to the phosphorylated β-catenin and β-catenin is then degraded by ubiquitin-mediated proteasomal degradation machinery. In the presence of Wnt ligand, it binds to the Frizzled/LRP5/6 receptor complex, leading to the dissociation of the destruction complex. Free β-catenin proteins accumulate in the cytosol and are then translocated into the nucleus for activation of TCFs/LEFs (T-cell factors/lymphoid enhancing factors). The activation of TCFs/LEFs transcription factors induces a variety of Wnt/β-catenin signaling target genes, including those involved in skin wound healing (e.g., *Axin2*, *Collagen I*, *Collagen III*, *EGFR*, *Endothelin-1*, *Fibronectin*, *Keratin-14*, *Lgr5*, *VEGF*, *WISP-1*).

Figure 4

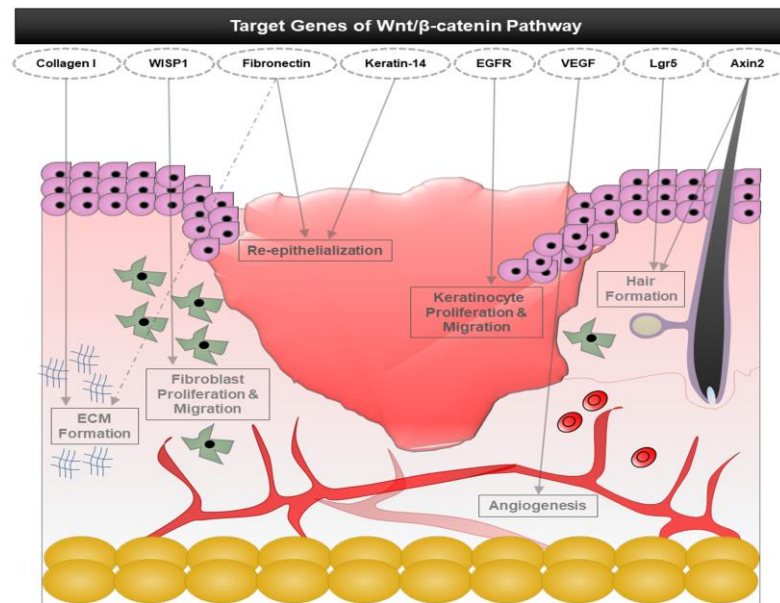


Figure 4. The effects of Wnt/β-catenin pathway target genes on cutaneous wound repair

Various Wnt/β-catenin signaling target genes contribute to the multiple events that occur during wound healing. For example, Collagen-I has a role in ECM formation, WISP1 activates fibroblast proliferation and migration, fibronectin controls re-epithelialization and ECM formation, Keratin-14 promotes re-epithelialization, EGFR regulates keratinocyte proliferation and migration, VEGF enhances angiogenesis, and Lgr5 and Axin2 promotes hair formation via activation of hair follicle stem cells.

Figure 5

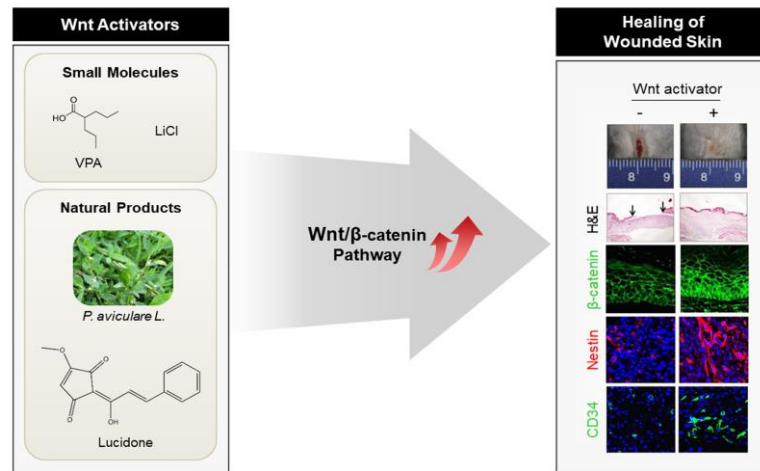


Figure 5. Effects of Wnt activators on healing of wounded skin

The agents that activate the Wnt/ β -catenin pathway include small molecules, such as LiCl and VPA (valproic acid), and natural products, such as lucidone and *P. aviculare* (*Polygonum aviculare L.*) extract.^{112,115} Up-regulation of the Wnt/ β -catenin pathway via topical application of these agents promotes wound healing with increased expression of stem cell markers such as nestin and CD34. Reproduced with permission from Lee et al. and Seo et al.^{112,115}

Figure 6

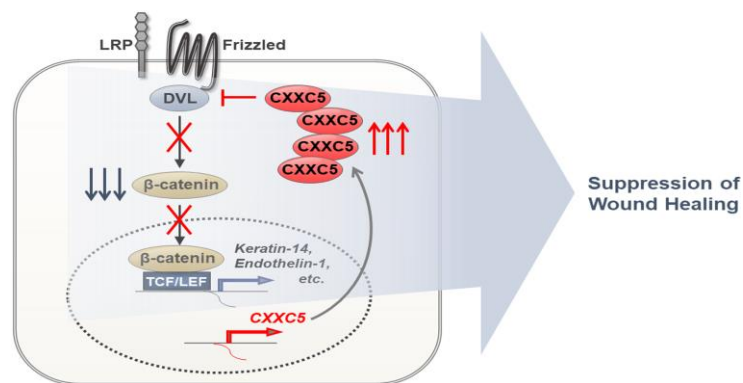


Figure 6. CXXC5 is a negative feedback regulator of the Wnt/ β -catenin pathway and suppresses expression of target genes involved in wound healing.

CXXC5 transcription is induced by strong activation of the Wnt/ β -catenin during the wound healing process. CXXC5 binds to DVL in the cytosol and subsequently suppresses Wnt/ β -catenin signaling by blocking the dissociation of the destruction complex by DVL. Inhibition of Wnt/ β -catenin signaling in the skin results in repression of wound healing-related genes following inhibition of the wound healing process.

Figure 7

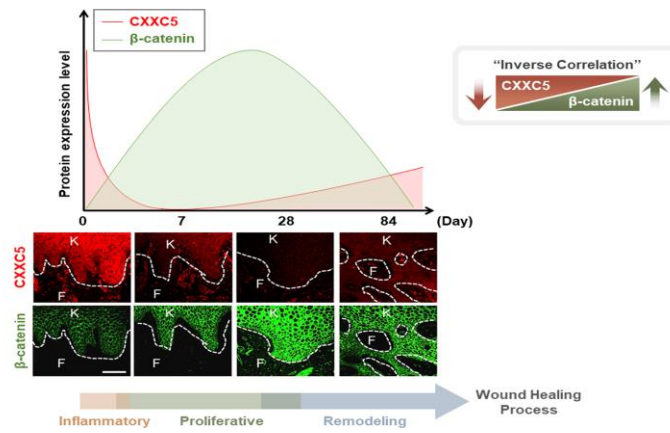


Figure 7. Kinetics of the expression levels of β -catenin and CXXC5 during the wound healing process

Profiles for CXXC5 and β -catenin expression during the wound healing process, adapted from a previous study.¹³ The tissue samples were from patients with melanoma who underwent surgery. The immunohistochemical images represent expression levels of CXXC5 and β -catenin in wounded skin at 0, 7, 28, and 84 days after surgery. Day 0, intact skin. White dashed lines, the epidermal-dermal junction. F, fibroblasts; K, keratinocytes. Scale bars, 100 μ m. Reproduced with permission from Lee et al.¹³

Figure 8

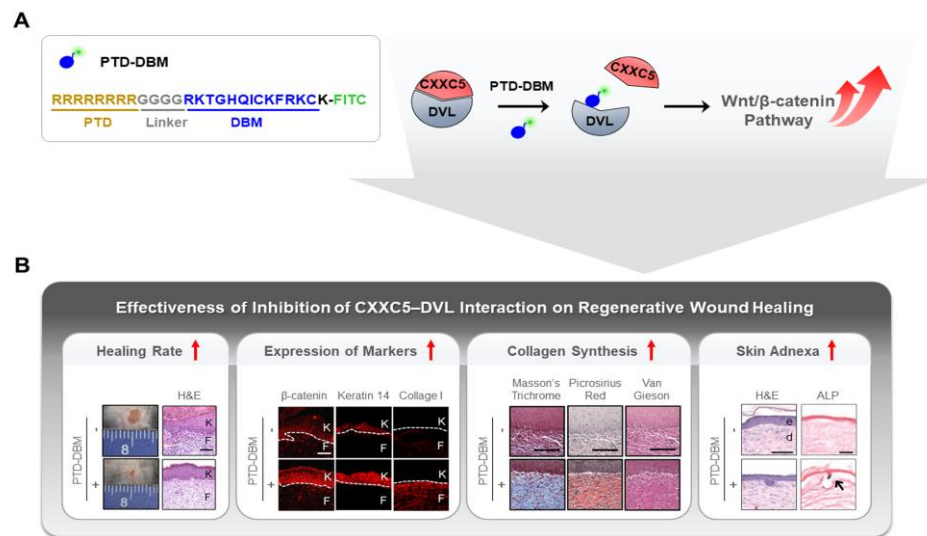


Figure 8. PT-D-DBM, a peptide that interferes with the CXXC5-DVL interaction, and its effectiveness on regenerative wound healing

(A) The PT-D-DBM peptide consists of a protein transduction domain (PTD), linker, DVL-binding motif (DBM), and FITC (left).⁸⁰ The function of the PT-D-DBM peptide is exerted by interfering with the CXXC5–DVL interaction via competitive DVL binding with CXXC5, followed by activation of the Wnt/β-catenin signaling (right). **(B)** The effects of PT-D-DBM on wound healing in mice. The immunohistochemical images are adapted from a previous study.^{13,124} 100 μM of PT-D-DBM was applied daily into the wounded skin of 7-week-old male C3H mice for 11 days after wound formation (diameter = 1.5 cm). Analysis of the healing rate is shown, macroscopic image and H&E staining results. Confocal microscopic examination was used to detect the expression of β-catenin, keratin-14, and collagen-I markers. Collagen synthesis was measured using Masson's trichrome, picrosirius red, and van Gieson staining. For analysis of skin adnexa, 2 mM of PT-D-DBM was applied daily into the wounded skin of 3-week-old male C3H mice for 14 days after wounding (diameter = 1 cm). The formation of neogenic hair follicles was detected using H&E and alkaline phosphatase (ALP) staining (dark blue). White dashed lines, the epidermal-dermal junction. Black arrow, ALP expression. F, fibroblasts; K, keratinocytes; e, epidermis; d, dermis. Scale bars, 100 μm. Reproduced by permission from Lee et al.^{13,124}

Advances in Wound Care
Approaches for regenerative healing of cutaneous wound with an emphasis on strategies activating the Wnt/ β -catenin pathway (DOI: 10.1089/wound.2020.1284)
This paper has been peer-reviewed and accepted for publication, but has yet to undergo copyediting and proof correction. The final published version may differ from this proof.



Table 1. List of target genes involved in the Wnt/ β -catenin pathway that are related to wound healing

Wnt target genes	Role in wound healing	Reference
Axin2	Hair formation via activation of hair follicle stem cells	77,126,127
Collagen I	Key protein of ECM synthesized during proliferative phase	6,104,128
Collagen III	Key protein of ECM synthesized during early proliferative phase	104,129,130
EGFR	Regulation of keratinocyte migration to wound bed	131
Endothelin-1	Regulation of fibrosis and calcification	132
Fibronectin	ECM formation and re-epithelialization	133,134
Keratin-14	Re-epithelialization	<u>135</u>
Lgr5	Hair formation via activation of hair follicle stem cells	<u>77,98</u>
VEGF	Stimulation of angiogenesis	136,137
WISP1	Promotion of dermal fibroblast proliferation and migration	138,139

Table 2. The effects of Wnt/ β -catenin pathway activation on wound repair and regeneration

	The effects by activation of Wnt/ β -catenin pathway	Reference
Repair	• Promotion of angiogenesis	90,140
	• Promotion of fibroblast migration, proliferation, and differentiation	89,141
	• Promotion of keratinocyte proliferation and differentiation	142,143
	• Stimulation of re-epithelization	24
	• Induction of ECM formation	87,144
Regeneration	• Stimulation of wound-induced hair folliculogenesis	22,127
	• Enhancement of epidermal stem cells proliferation and differentiation	93,94
	• Induction of hyaluronic acid synthesis	101

ECM, extracellular matrix

Table 3. The Wnt/ β -catenin pathway activators enhancing wound healing

Agents for wound healing	Type	Target Wnt component	Reference
LiCl	Small molecule	Inhibition of GSK3 β	94
VPA	Small molecule	Inhibition of GSK3 β	112
Lucidone	Natural product	Inhibition of GSK3 β	113
<i>Polygonum aviculare L.</i>	Natural product	Various targets	115
PTD-DBM	Peptide	Inhibition of CXXC5–DVL interaction	13,124
Wnt3a	Recombinant protein	Wnt ligand	101

LiCl, Lithium chloride; VPA, valproic acid; PTD, protein transduction domain; DBM, DVL binding motif